Genomeditech (Shanghai) Co.,Ltd.
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Product Sheet

Luciferase OVCAR3 Cell Line

Catalog number: GM-C31422

Version 3.3.1.250611

Description	Luciferase OVCAR3 Cell Line is a clonal stable OVCAR3 cell line that constitutively expresses the Luciferase gene, constructed using lentiviral technology.
Quantity	4E6 Cells per vial,1 mL
Product Format	3 vials of frozen cells
Shipping	Shipped on dry ice
Storage Conditions	Liquid nitrogen immediately upon receipt

Target	/
Gene ID/Uniprot ID	/
Host Cell	OVCAR3

Recovery Medium	RPMI 1640(ATCC)+20% FBS+1% P.S+0.01 mg/mL Bovine Insulin
Growth medium	RPMI 1640(ATCC)+20% FBS+1% P.S+0.01 mg/mL Bovine Insulin+0.25 μg/mL Puromycin
Note	Cells should be cultured using ATCC/30-2001 RPMI 1640 medium or Growth medium from Genomeditech. The serum should be Cegrogen biotech/A0500-3010 or sourced from Gibco.
Freezing Medium	90% FBS+10% DMSO
Growth properties	Adherent
Growth Conditions	37°C, 5% CO_2

Mycoplasma Testing	The cell line has been screened to confirm the absence of Mycoplasma species.
Safety considerations	Biosafety Level 2
Note	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

Materials

Reagent	Manufacturer/Catalogue No.
RPMI 1640	gibco/C11875500BT
Fetal Bovine Serum	Cegrogen biotech/A0500-3010
Pen/Strep	Thermo/15140-122
Puromycin	Genomeditech/GM-040401
96 well White Flat Bottom Polystyrene Not Treated Microplate	Corning/3912
GMOne-Step Luciferase Reporter Gene Assay Kit	Genomeditech/GM-040503

Figures

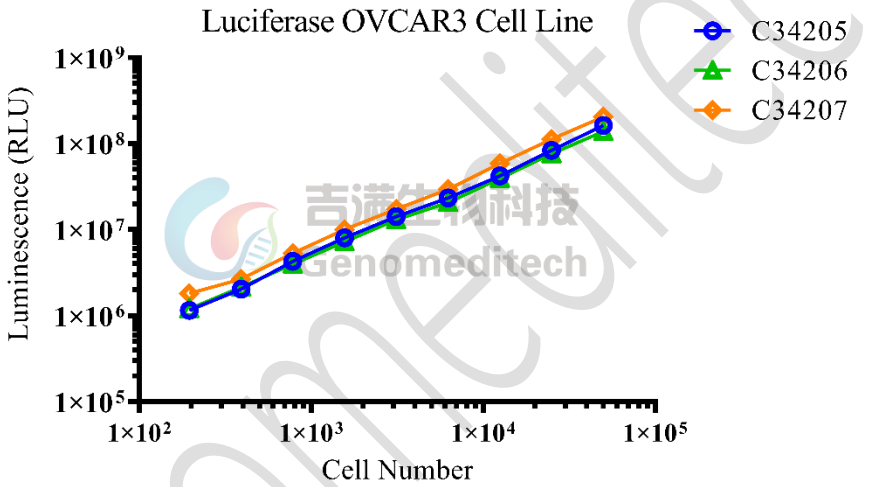

Figure 1 | Correlation between the number of cells and bioluminescence values. Serial dilutions of Luciferase OVCAR3 Cell Line (Cat. GM-C31422) (96-well format). The firefly luciferase activity was measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. GM-040503).

Cell Recovery

Recovery Medium: RPMI 1640(ATCC)+20% FBS+1% P.S+0.01 mg/mL Bovine Insulin

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

a) Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).

b) Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.

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c) Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium and spin at approximately 176 x g for 5 minutes. Discard supernatant.

d) Resuspend cell pellet with the recommended recovery medium. And dispense into appropriate culture dishes.

e) Incubate the culture at 37°C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

a) Centrifuge at 176 x g for 3 minutes to collect cells.

b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 4E6 cells/mL.

c) Aliquot 1 mL into each vial.

d) Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: RPMI 1640(ATCC)+20% FBS+1% P.S+0.01 mg/mL Bovine Insulin+0.25 μg/mL Puromycin

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

a) Subculturing is necessary when the cell density reaches 80%. It is recommended to perform subculturing at a ratio of 1:2 to 1:3 every 3 to 4 days.

b) Remove and discard culture medium.

c) Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.

d) Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 3 to 4 minutes at 37°C).

e) Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.

f) Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.

g) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.

h) Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:2 - 1:3 is recommended

Medium Renewal: Every 3 to 4 days

Notes

a) Cell growth rate is slow. Repeated pipetting during digestion should be avoided, as it may result in severe cell aggregation. Cell morphology is heterogeneous, with some large cells containing vacuoles. The presence of black granules within the cytoplasm of these cells is considered a normal phenomenon.

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Labeled Cells	
Luciferase-GFP MCF-7 Cell Line	Luciferase A498 Cell Line
Luciferase B16-F10 Cell Line	Luciferase HL-60 Cell Line
Luciferase MIA PaCa-2 Cell Line	Luciferase MM.1R Cell Line
Luciferase NCI-H929 Cell Line	Luciferase U-937 Cell Line
Luciferase-ZsGreen1 K562 Cell Line	Luciferase-ZsGreen1 Raji Cell Line
D-Luciferin, Potassium Salt	D-Luciferin, Sodium Salt

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